

Triatomines: Trypanosomatids, Bacteria, and Viruses Potential Vectors?

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Vieira CB, Praça YR, Bentes KLS, Santiago PB, Silva SMM, Silva GS, Motta FN, Bastos IMD, Santana JM and Araújo CN (2018) Triatomines: Trypanosomatids, Bacteria, and Viruses Potential Vectors? Front. Cell. Infect. Microbiol. 8:405. doi: 10.3389/fcimb.2018.00405 Triatominae bugs are the vectors of Chagas disease, a major concern to public health especially in Latin America, where vector-borne Chagas disease has undergone resurgence due mainly to diminished triatomine control in many endemic municipalities. Although the majority of Triatominae species occurs in the Americas, species belonging to the genus Linshcosteus occur in India, and species belonging to the Triatoma rubrofasciata complex have been also identified in Africa, the Middle East, South-East Asia, and in the Western Pacific. Not all of Triatominae species have been found to be infected with Trypanosoma cruzi, but the possibility of establishing vector transmission to areas where Chagas disease was previously non-endemic has increased with global population mobility. Additionally, the worldwide distribution of triatomines is concerning, as they are able to enter in contact and harbor other pathogens, leading us to wonder if they would have competence and capacity to transmit them to humans during the bite or after successful blood feeding, spreading other infectious diseases. In this review, we searched the literature for infectious agents transmitted to humans by Triatominae. There are reports suggesting that triatomines may be competent vectors for pathogens such as Serratia marcescens, Bartonella, and Mycobacterium leprae, and that triatomine infection with other microrganisms may interfere with triatomine-T. cruzi interactions, altering their competence and possibly their capacity to transmit Chagas disease.

Keywords: kissing bugs, vector competence, infectious diseases, Chagas disease, pathogens

INTRODUCTION

Vector-borne diseases (VBDs) are human illnesses caused by parasites, viruses, and bacteria that are usually transmitted by a bloodsucking arthropod. According to the World Health Organization (WHO) VBDs together account for around 17% of all infectious diseases in the world, which are mainly distributed in tropical and subtropical areas, affecting poorest populations. However, environmental changes and globalization have placed many more people living in other parts of the world at risk of contracting VBDs. The emergence/re-emergence of these diseases is burdening the health systems in many countries due to possible epidemic outbreaks and their impact in morbidity and mortality (WHO, 2017).

Triatominae insects (Hemiptera: Reduviidae) (Lent and Wygodzinsky, 1979; Jurberg et al., 2014) are a diverse subfamily of ectoparasites mainly distributed across Americas (Guarneri et al., 2000; Otálora-Luna et al., 2015) (Figure 1). Six species belonging to the genus Linshcosteus occur in India, and species belonging to Triatoma have been also identified in Africa, the Middle East, South-East Asia and in the Western Pacific (Figure 1, Table S1). Currently, the 151 Triatominae species described are grouped in 17 genera and organized into 5 tribes: Aberproseniini, Bolboderini, Cavernicolini, Rhodiniini, and Triatomini (Table S1) (Alevi et al., 2014, 2016; Souza et al., 2016; da Rosa et al., 2017). They are typically found in a wide range of sylvatic environments frequently associated to several wild vertebrate hosts, possibly in their nests and burrows. Contact with humans occurs when men enter in forested areas or when they reach urban areas by flying and colonize human dwellings (Lazzari et al., 2013).

All species are considered able to transmit *Trypanosoma cruzi*, agent of Chagas disease. However those belonging to the genera *Triatoma*, *Rhodnius* and *Panstrongylus* have greatest epidemiological importance in Latin America (Cavassin et al., 2014), as they have adapted to colonize peridomestic and domestic environments, coming into close contact with domestic animals and humans. Domiciliation occurs through the loss of habitat due to irregular environment exploitation, such as deforestation and burnings (Dias and Schofield, 1998; Almeida et al., 2009) or through the loss of primary hosts, which can triggers a switch to accessible humans (Schofield, 1988).

T. cruzi and other pathogens infection within Triatominae colonies is due mainly to the vector blood feeding behavior. They are obligate hematophagous insects, exhibiting behavioral, morphological and molecular adaptations to feed on a variety of vertebrate blood throughout their life cycle (Lazzari et al., 2013), in which they undergo five nymphal stages before reaching adulthood (Jurberg et al., 2014). Therefore, their dietary behavior ensures the scenario to act as potential vectors of multiple pathogens to humans at any developmental stage. Moreover, exposure to triatomine bites and feces can result in immunological reactions (Walter et al., 2012), such as swelling, severe fever, itching at the bite site, and anaphylaxis (Klotz et al., 2010; Dujardin et al., 2015).

Thus, from the close relationship of Triatominae with vertebrate hosts, beyond T. cruzi, different triads of interaction (vector/pathogen/vertebrate host) may arise, allowing new infection routes for other microorganisms, such as bacteria and viruses, in cases Triatominae species show vector competence and vectorial capacity. Vectorial capacity is the ability of a vector to transmit a pathogen in a given locality at a definite time. It comprises the vector interactions with the infectious agent and with the vertebrate host(s), and is affected, for instance, by vector density, longevity, host preference, feeding behavior, and vector competence. Precisely, vector competence embodies vectorpathogen interactions, encompassing specifically the ability of a vector to become infected by and transmit a pathogen. It includes susceptibility to infection, permissiveness for pathogen reproduction and development, transmission efficiency and duration of extrinsic incubation period, which is the time from the uptake of an infectious meal to the time it is capable of transmitting the pathogen (Higgs and Beaty, 2004).

To be a competent vector, triatomine has to acquire the pathogenic microorganism from an infected host, to allow its replication in the midgut or to spread to the hemocoel in order to enter into the salivary glands. Hence, pathogens may be carried from the bug to the human host during insect bite by two different routes: via contaminated saliva released into host blood vessel or via contact with contaminated feces deposited at the host skin or near mucosa. It is worth mentioning that oral infection may also occur in case of food contamination with triatomine feces/saliva. An additional route of pathogen transmission may be xenodiagnosis, a diagnostic method that uses Triatominae nymphs as a biological culture medium for the detection of T. cruzi infection. Pathogens have been developing adaptations to exploit vector biology, behavior and ecology, and mainly, different mechanisms of transfer from one host to another to continue its species maintenance. Understanding all possible spreading routes of pathogens is crucial to prevent infectious diseases. Thus, considering the diversity of pathogens that Triatominae may acquire during blood feeding, the aim of this review was to examine published literature and summarize current information of triatomine competence to transmit, beyond T. cruzi, other pathogens to humans.

MICROORGANISMS TRANSMITTED THROUGH TRIATOMINE/MAN CONTACT AND THEIR POSSIBLE ROLE IN HUMAN HEALTH

Trypanosomatids

Trypanosoma cruzi and Chagas Disease

The Trypanosomatidae family encompasses various obligate parasites, including the protozoa T. cruzi that causes Chagas disease (Hoare, 1972; Cavalier-Smith, 2016). The transmission of Chagas disease by Triatominae is very well reported in literature. Infection with vector-borne T. cruzi begins when metacyclic trypomastigotes, which are motile forms of the parasite, penetrate into the vertebrate host through the triatomine feces and urine. Once in the vertebrate host, these forms, which have evolved to survive inside host cells, infect nucleated cells. Within the cell, they differentiate into amastigotes in a phagosomal compartment known as the parasitophorous vacuole, escape to the cytoplasm and replicate asexually through longitudinal binary division to form several amastigotes. As the cell becomes full of amastigotes, these convert into trypomastigotes and breach it, invading adjacent tissues and spreading to distant sites through bloodstream and lymphatics. The parasite population expands due to repeated cycles of cell invasion and replication, which lead to immune responses and can give rise to Chagas-associated pathologies (Tyler and Engman, 2001).

When triatomines take a blood meal, bloodstream trypomastigotes enter the midgut and differentiate into epimastigotes, amastigotes, and spheromastigotes (Rassi et al., 2010; Teixeira et al., 2011). Epimastigotes divide through



longitudinal binary asexual reproduction repeatedly, which then migrate to the rectum, where they differentiate into infective and non-replicative metacyclic trypomastigotes, process called metacyclogenesis (Figure 2). T. cruzi infection intensity is triatomine species specific (Kollien and Schaub, 2000), as the conditions for its development in the gut vary substantially (Garcia et al., 2010). Therefore, the protozoan is capable of causing significant physiological changes such as delay in the development of Triatoma infestans nymphs (Schaub, 1989), decline in Rhodnius prolixus adult life span (Schaub, 1989) and of blood meal ingestion in infected Triatoma dimidiata nymphs (Schaub et al., 2011). Although epimastigotes and metacyclic trypomastigotes are eliminated along with the insect feces, only the latter are able to infect vertebrates (Schuster and Schaub, 2000), reach the vertebrate bloodstream through preexistent lesions or bite resultant breaches of the skin, or through mucosal tissues, repeating the life cycle (Billingsley and Downe, 1986; Figueiredo et al., 2000; Garcia et al., 2007). A balance among T. cruzi strain, inoculated burden, the innate and acquired immunological responses seems to be critical for the parasite control in the vertebrate host (Tarleton, 2007; Gil-Jaramillo et al., 2016).

Endemic to the continental part of Latin America, Chagas disease (American trypanosomiasis) affects approximately 8 million people worldwide (WHO, 2018a). It has been increasingly detected in non-endemic countries like USA, Canada, and countries in Europe and Western Pacific Region, due primarily to people movement between Latin America and the rest of the world, being an emerging infectious disease in these regions (WHO, 2018b). It is interesting to note

that in the southern of USA recent surveillance data unveil Triatominae adults and nymphs in domestic and peridomestic areas, increasing human *T. cruzi* infection by vector-borne route, becoming Chagas disease a public health concern in this country (Montgomery et al., 2014; Edwards et al., 2017; Curtis-Robles et al., 2018).

In Vietnam, in the southeastern Asia, *Triatoma rubrofasciata* has been reported in high densities associated with rats and chickens close to human dwellings in rural and, specially, in urban areas. In the latter, *T. rubrofasciata* adults were reported hiding in people beds and feeding on them while sleeping, in buildings from the ground to the eighth floor. Although it is known this species transmit *T. cruzi* in Latin America and *Trypanosoma conorhini* worldwide, these trypanosomatids have not been detected in blood smears from individuals bitten by this bug. Closely related to *T. rubrofasciata* complex, *Linshcosteus* is uniquely distributed in Indian. Due to human migration and triatominae anthropophilia, *T. cruzi* infection emergence in Asia and India can arise (Dujardin et al., 2015).

Chagas disease warrants attention as it is considered the parasitic disease with the major socioeconomic impact in Latin America, being responsible for loss of productivity with an estimated value of US\$ 1.2 billion annually (WHO/TDR, 2012).

Trypanosoma rangeli

Another parasite from the Trypanosomatidae family infecting humans and other vertebrates is *T. rangeli* (D'Alessandro, 1976; Guhl and Vallejo, 2003). Five phylogenetic lineages named TrA, TrB, TrC, TrD, and TrE have been identified to date, TrB being the most phylogenetically divergent one (Da Silva et al., 2004,



2009; Maia da Silva et al., 2004; Maia Da Silva et al., 2007; Ortiz et al., 2009; Caballero et al., 2015). TrA, TrB, and TrC lineages were reported to infect humans (Da Silva et al., 2004, 2009; Maia Da Silva et al., 2007; Ortiz et al., 2009; Pinto et al., 2012, 2015; Sincero et al., 2015; Dario et al., 2016, 2017). *T. rangeli* differs in life cycle, host infection and immune evasion both in triatomine and vertebrate hosts in comparison with *T. cruzi* (Espinosa-Álvarez et al., 2018).

Vector-borne T. rangeli transmission begins when metacyclic trypomastigotes penetrate into the vertebrate host through the triatomine saliva. Paim et al. (2013) showed that a 5th nymph of a T. rangeli-infected R. prolixus could carry up to 100,000 metacyclic trypomastigotes in its salivary glands. Approximately half of them can be released during blood feeding into the bite site (Ferreira Lde et al., 2015). Once in the vertebrate hosts, these forms may develop a multiplicative cycle. There is no strong evidence of reproductive forms of T. rangeli in these hosts, although some studies suggest their presence in vertebrates (de Scorza et al., 1986; Urdaneta-Morales and Tejero, 1986; Osorio et al., 1995; Ferreira Lde et al., 2015). Moreover, Ferreira et al. suggest that T. rangeli can hide in some tissues or organs, as it was rarely found circulating in mouse blood, even after an infected triatomine had injected a huge number of parasites (Ferreira Lde et al., 2015).

T. rangeli (sub-genera *Tejeraia*), as well as *Trypanosoma brucei* and *Trypansoma evansi* (sub-genera *Trypanozoon*), is classified as a Salivarian trypanosome because it multiplies in the anterior part of the vector digestive tract (Desquesnes et al., 2007). So, when triatomines feed on *T. rangeli*-infected hosts, trypomastigotes get into the midgut and differentiate into epimastigotes. These forms cross the gut epithelium, replicate in the hemolymph

and then reach the salivary glands where they differentiate into metacyclic trypomastigotes. The cycle reinitiates when these forms are transmitted to mammals during the next triatomine bite (D'Alessandro, 1976) (**Figure 2**). *T. rangeli* is transmitted specially by Triatominae of the Rhodniini tribe. Its metacyclic trypomastigotes were already reported in *Rhodnius domesticus*, *Rhodnius nasutus*, *Rhodnius neglectus*, *Rhodnius pallescens*, *R. prolixus*, *Rhodnius robustus*, *Rhodnius brethesi*, *Rhodnius colombiensis*, *Rhodnius ecuadoriensis*, *Rhodnius dalessandroi*, *Rhodnius pictipes*, and *Rhodnius neivai* salivary glands (Guhl and Vallejo, 2003; Vallejo et al., 2009, 2015; Castro et al., 2017). Nevertheless, it was also reported in *Triatoma carrion*, *Panstrongylus chinai* and *Panstrongylus rufotuberculatus* species from the Triatominii tribe (Ocaña-Mayorga et al., 2015).

T. rangeli is capable of causing reduction in *R. prolixus* fecundity and fertility (Fellet et al., 2014), reduction in negative phototaxis and in *R. prolixus* expression of cGMP-dependent protein kinase gene, an ortholog of *Drosophila melanogaster foraging* gene reported to control the locomotion activity (Marliére et al., 2015), increase in mortality, defective, delayed or absent molts and tissue damages (Ferreira et al., 2010), among other effects in the infected triatomines (Guhl and Vallejo, 2003; Peterson and Graham, 2016). However, it seems *R. prolixus* co-infection with *T. rangeli* and *T. cruzi* ameliorate adverse effects of distinct infections, helping the insect to tolerate greater parasite infections (Peterson et al., 2016).

T. rangeli occurs from Central to South America (Grisard et al., 1999b). Although it is not considered a human pathogen, this trypanosomatid deserves attention as it shares biological characteristics, antigens, geographical distribution, insect and vertebrate hosts with *T. cruzi*. Mixed infections with *T. cruzi*

are observed in humans, sylvatic and domestic mammals (Grisard et al., 1999b; Guhl and Vallejo, 2003). Thus, T. rangeli transmission by T. cruzi vectors is biologically and epidemiologically important due to crossed serological reactions (Guhl and Marinkelle, 1982; Guhl et al., 1987; Coura et al., 1996; Cuba Cuba, 1998; Calzada et al., 2010), what may result first in increased diagnostic test costs, and second in harms to human health due to Chagas disease drug toxicity where false positives are not detected (Grisard et al., 1999a). To circumvent this possibility, Moraes et al. evaluated the serological cross-reactivity between epimastigotes and trypomastigotes from T. cruzi and T. rangeli using sera of acute and chronic chagasic patients by indirect immunofluorescence and immunoblotting assays. These authors recommend retesting, in areas where there is overlapped distribution of both species, Chagas disease positive sera with T. rangeli trypomastigote antigens to avoid misdiagnosis (de Moraes et al., 2008).

Bacteria

Serratia marcescens

Serratia genus (Enterobacteriaceae family) contains 14 species and 2 subspecies (Mahlen, 2011). They have been associated with human infections and reported as a symbiont in the microbiota of hematophagous insects (Grimont and Grimont, 1978; Grimont et al., 1979; Azambuja et al., 2004). In humans, S. marcescens is an important opportunistic gram-negative bacteria reported to cause wound, urinary tract, bloodstream and ocular infections, pneumonia and other respiratory diseases, meningitis, endocarditis, among other clinical diseases (Mahlen, 2011; González-Juarbe et al., 2015). The clinical and epidemiological significance are associated with sporadic hospital infection outbreaks due to S. marcescens strains ability to produce βlactamases, which confer resistance to β-lactams and antiseptic agents available (Maragakis et al., 2008; Carvalho et al., 2010). Many outbreaks are frequently reported worldwide especially in neonatal intensive care units, where the bacteria usually spread rapidly and is associated with significant morbidity and mortality. The sources of the outbreaks have been associated to contaminated laryngoscope blades, hands, ventilator equipment, disinfectants, and breast pumps (Gransden et al., 1986; Jones et al., 2000; Sartor et al., 2000; Jang et al., 2001; Villari et al., 2001; Fleisch et al., 2002; Uduman et al., 2002; Cullen et al., 2005; Montagnani et al., 2015; Åttman et al., 2018).

Azambuja et al., isolated *S. marcescens* biotype A1a from the midgut of labroratory-reared *R. prolixus* (Azambuja et al., 2004), confirming previous reports on the occurrence of this species in triatomine gut (Figueiredo, 1995). Following feeding, this gramnegative bacillus rapidly multiplied and showed hemolytic and trypanolytic effects, the latter on *T. cruzi* Y strain but not on *T. cruzi* DM28c clone. Many strains of *S. marcescens* produce a reddish pigment named prodigiosin that is an important compound for the action against the parasite in the insect midgut (Azambuja et al., 2004). This report provides evidence that *R. prolixus* may be a competent vector for *S. marcescens*, as it was able to maintain infection in the gut, and that bacteria may be eliminated within the feces. Also suggests that *S. marcescens* may compete with *T. cruzi*, reducing *R. prolixus* competence for this

trypanosomatid. On the other hand, *S. marcescens* biotype A1a did not raise triatomine mortality (Azambuja et al., 2004). The conduction of transmission studies from triatomines to mice would help to confirm the potential of *R. prolixus* to serve as vector for *S. marcescens*.

To date *S. marcescens* vector-borne transmission was not reported, but it is important to emphasize this possible mode of transmission. It is possible *S. marcescens* living in triatomine midgut to be acquired during the consumption of food containing the contaminated feces of an infected triatomine or maybe vector-borne through contaminated feces contact with open wounds or mucous membranes of a susceptible host. Future studies may evidence this possibility and indicate the consequences for human health.

Bartonella

Bartonella genus belongs to the Bartonellaceae family of the alphaproteobacteria (Regier et al., 2016), along with *Rickettsia* and *Brucella* (Sanchez Clemente et al., 2012) and contains 36 species, characterized by coccobacillary or bacillary rods. These pleomorphic fastidious gram-negative bacteria are considered a facultative intracellular pathogen (Maguiña and Gotuzzo, 2000). Human body lice, sandflies, cat fleas, flies and ticks are *Bartonella* vectors to humans and other mammals (Tsai et al., 2011). The vector-borne is the most relevant route of transmission (Pons et al., 2016).

A novel Bartonella genotype, named Candidatus Bartonella rondoniensis, closely related to Bartonella bacilliformis and Bartonella ancashensis, severe human pathogens, was described in the sylvatic triatomine Eratyrus mucronatus (Laroche et al., 2017). This new genotype is also related to B. bovis that has been observed to cause endocarditis in cats, which are accidental hosts of this species (Maillard et al., 2007). E. mucronatus feeds on bats and other small mammals (Castro et al., 2010), which are sources of virus and bacteria (Mühldorfer, 2013), including Bartonella (Laroche et al., 2017). Although sylvatic, this triatomine is attracted to artificial light sources (Castro et al., 2010) and has been reported in the proximity and inside houses. By now, Laroche et al. suggest that this species may host pathogenic bacteria (Laroche et al., 2017). An experimental model of infection would help to confirm if E. mucronatus is a competent vector for Bartonella species and act as their vector to humans. One possibility is that in these arthropods, Bartonella migrates to, replicate in the salivary glands and is transmitted during the bite.

In humans, *B. bacilliformis* is transmitted through the bite of infected *Lutzomyia verrucarum*, *Lutzomyia peruensis*, among other phlebotomine sandflies. This disease, named Carrion's disease, is endemic in South America and was the first human bartonellosis described. Carrion's disease can occur in two distinct concomitant or independent syndromes. The first is known as Oroya fever, which is characterized by hemolytic fever and bacteremia that without treatment can cause 40–88% death. Verruga peruana is the second syndrome, characterized by hemangiomas owed to endothelium infection (Minnick et al., 2014). *B. bacilliformis* closely related *B. ancashensis* was isolated from patients' blood with chronic verruga peruana, being suggested as a second agent of the disease (Mullins et al., 2015). Other *Bartonella* species may cause endocarditis, *B. quintana* causes trench fever and *B. henselae* causes cat scratch disease (Regier et al., 2016).

Mycobacterium leprae

M. leprae belongs to Mycobacteriaceae family and causes leprosy, a disfiguring chronic systemic infectious disease. Early diagnosis and the available treatment with multi-drug therapy significantly reduced the disease burden in recent decades. However, the lack of awareness about early signs of leprosy that contributes to a delay in diagnosis, and the persistent stigma and discrimination against affected people are factors that complicate the fight against leprosy. The global incidence of new cases was approximately 213,000 in 2014, and the highest prevalence rates are observed in Brazil, India and Indonesia, which together account for 81% of the newly diagnosed cases globally (WHO, 2016). To date, the exact mechanism of leprosy transmission is not completely understood, but it is supposed that transmission occurs due to the inhalation of infectious aerosols released by untreated cases of the disease or by direct contact from an infected person to a susceptible individual (Scollard et al., 2006; WHO, 2015). Interestingly, studies point out the viability of M. leprae outside human body and its existence in the environment, suggesting a different possibility in disease transmission (Desikan and Sreevatsa, 1995; Matsuoka et al., 1999; Turankar et al., 2012, 2016).

Recently, a report from Triatominae gut microbiomes revealed the presence of Mycobacterium in Triatoma protracta species microbiome profile (Rodríguez-Ruano et al., 2018). A further study evaluated the potential of M. leprae transmission by some insects and it was shown that *R. prolixus* might be able to transmit these bacteria, once they were present in R. prolixus feces (Neumann Ada et al., 2016). Perhaps the approximately 50% of Triatominae species found in Brazil, Linshcosteus (6 species) in India and T. rubrofasciata in Indonesia (Table S1) could help leprosy transmission in these three countries with the highest global incidences of the disease. McFadzean and Macdonald evaluated the possible role of mosquitoes and bed bugs in leprosy transmission by allowing infected and control insects to take a blood meal on volunteers and found no difference in transmission (McFadzean and Macdonald, 1961). Almost two decades later, the presence of *M. leprae* was reported in the proboscis, cuticle and blood smears from mosquitoes and ticks (de Souza-Araujo, 1942; Banerjee et al., 1991), suggesting arthropods could act as biological or mechanical vectors to these bacilli (Kirchheimer, 1976; Benchimol and Romero Sa, 2003).

Armadillos (*Dasypus novemcinctus*), which are parasitized by triatomines (Lainson et al., 1979), are a natural reservoir for *M. leprae* (Walsh et al., 1975; Smith et al., 1983) and also for *T. cruzi* (Lainson et al., 1979; Paige et al., 2002). Moreover, it is important to highlight that a study reported wild armadillos as well as patients with leprosy infected with the same strain of *M. leprae* (Truman et al., 2011). In this context, it is a topic of concern the overlapping geographic distribution of Triatominae species and the endemicity of leprosy in some regions, as triatomines that are transitioning from wild environments to the domiciliary ones

may be a potential source of *M. leprae* transmission to humans (Neumann Ada et al., 2016), supporting that old hypothesis that leprosy can be vector-borne transmitted through the insect feces containing the bacteria when in contact with host wound or mucosa (Kirchheimer, 1976; Benchimol and Romero Sa, 2003; Neumann Ada et al., 2016).

Virus

Human Immunodeficiency Virus (HIV) and Hepatitis B Virus (HBV)

HIV (genus *Lentivirus*, *Retroviridae* family) is own singlestranded RNA virus that infects human CD4⁺ T lymphocytes and macrophages. HIV genome contains nine ORFs that encode for 15 proteins (Frankel and Young, 1998), fundamental to replication and evasion from host defense. HIV causes infection in approximately 36.7 million people worldwide and was responsible for 1.0 million HIV-related causes deaths in 2016 (WHO, 2018c).

HBV (*Hepadnaviridae* family) infection causes acute and chronic hepatitis, and in severe forms, can lead to the development of cirrhosis and hepatocellular carcinoma (Liang, 2009). HBV causes infection in approximately 257 million people and was responsible for 887,000 deaths in 2015 (WHO, 2018d).

HIV- or HBV-infected patients were submitted to xenodiagnosis to investigate HIV or HBV possible transmission by triatomines (Granato et al., 1987; Nuzzo et al., 1998). For triatomines fed on AIDS patients in whose blood p24 HIV antigen was detected, no antigen was detected in the feces samples analyzed 24 h or 48 h post-blood meal (Nuzzo et al., 1998). For HBV, samples of feces and hemolymph were analyzed periodically and revealed that the hepatitis B surface antigen (HBsAg) was eliminated until 15 days post-blood meal. These authors concluded that although the HBV transit by the triatomine gastrointestinal tract, these insects may not yield efficient HBV infection (Granato et al., 1987).

In a previous study, human blood samples obtained from wild caught *Panstrongylus, Rhodnius*, and *Triatoma* were evaluated for the presence of HBsAg by haemagglutination and radioimmunoassay techniques. By the former, seven samples were presumably positive for HBsAg, but by the latter, only one, obtained from a *Triatoma sordida* specimen, was positive for the antigen (Candeias et al., 1976). These studies did not reveal a trend in viruses' transmission by triatomines, but instead highlight that efforts should be made to ensure greater consistence among studies that evaluate this task.

Triatoma Virus

Non-enveloped *Triatoma virus* (TrV) was first identified in *T. infestans* species in Argentina (Muscio et al., 1987). TrV belongs to the genus *Triatovirus* (Agirre et al., 2011) of the *Dicistroviridae* family (Czibener et al., 2000), in which viruses own positive-sense single-stranded RNA. Its genome contains two open reading frames, separated by an intergenic region, coding structural, and non-structural polyproteins (Johnson and Christian, 1998; Sasaki et al., 1998;



Czibener et al., 2000; Domier et al., 2000; Wilson et al., 2000).

So far, TrV is the only pathogenic virus described in Triatominae. TrV transmission among triatomines occurs vertically, through cannibalism or fecal-oral route (Muscio et al., 1987, 1988, 2000). It replicates in the intestinal epithelium cells of triatomines, causing leg paralyzes, delayed development, reduced fertility and death (Muscio et al., 1987, 1988; Rozas-Dennis and Cazzaniga, 2000).

TrV average prevalence in *T. infestans* is greater than of *T. cruzi* (Marti et al., 2009) and other microorganisms associated with triatomines (Muscio et al., 1997; Marti et al., 2009). Consequently, it has been considered that people living in Chagas disease endemic areas may have entered in contact with TrV through virus particles present in triatomine feces (Muscio et al., 2000). This supposition has been confirmed by the positive serology for anti-TrV antibodies in mice (Querido et al., 2013), hens (Muscio et al., 2000), and rabbits (Muscio et al., 1997) used to feed triatomines infected with TrV. Humans who live in endemic areas also showed positive TrV serology, among them 12.2% corresponded to healthy individuals (Querido et al., 2015), thus supporting the observation that TrV infection levels are higher/greater than *T. cruzi* infection levels in triatomines.

So far, the cases of TrV infection in vertebrates indicate that this virus is unable to replicate inside their cells (Muscio et al., 2000; Querido et al., 2013, 2015), and so may have no impact on human health. However, TrV may increase, indirectly, *T. cruzi* transmission to vertebrate hosts, strengthening triatomine vector competence for this trypanosomatid, although more studies are needed to confirm this hypothesis (Marti et al., 2017).

FINAL CONSIDERATIONS

In recent years, triatomines have undergone a major resurgence in the number of infestations, leading to re-emergence of *T. cruzi* vector transmission. Although the majority of Triatominae species occurs in the Americas, *Linshcosteus* spp. occur in India and *T. rubrofasciata* complex occurs in Africa, the Middle East, South-East Asia, and the Western Pacific countries (Dujardin et al., 2015). Although not all of Triatominae species are naturally infected with *T. cruzi*, there is the possibility of establishing vector transmission to areas where Chagas disease was previously non-endemic due to global population mobility. Furthermore, the resurgence and worldwide distribution of triatomines is concerning, considering they can enter in contact with other pathogens.

The possibility of these insects acting as vectors of other human pathogens aroused our curiosity. Triatomines are uncommon disease vectors since, except for salivarian trypanosomes as T. rangeli, pathogens seem unable to reach triatomine salivary glands in order to be transmitted by bite. Thus, the common route of transmission, as observed for T. cruzi, a stercorarian trypanosome, may be via contaminated feces released on skin wound or mucosa (Figure 3). We present reports of triatomine infected with vector pathogens (T. rangeli and TrV) already known to be transmitted to humans by triatomines, and of triatomines infected with human pathogens besides T. cruzi (the bacteria S. marcescens, Bartonella and M. leprae; and HIV and HBV) (Figure 3). Although no human viral disease can be attributed to Triatominae, TrV infection in triatomines may interfere with vector competence, augmenting the ability of triatomines to transmit Chagas disease.

Many factors may influence triatomine vector competence for transmitting pathogens to humans. The different regions in the gut themselves can bear unfavorable conditions, for instance pH, oxygen content, digestive enzymes, immune-related molecules, peristalsis, symbionts, and transient microorganisms (Douglas, 2015). Being competent does not mean to have the capacity to disseminate diseases, as vector competence is only one factor influencing vectorial capacity (Higgs and Beaty, 2004). Thus, further investigation and establishment of adapted models need to be developed in order to broaden the knowledge of triatominae ability to act as vector of other human infectious diseases.

AUTHOR CONTRIBUTIONS

CdA, CV, YP, and KB were involved in the conception of and wrote the manuscript. PS, SS, GS, FM, IB and JdS wrote the manuscript. PS, YP, and KB prepared the figures and tables. All the authors read and approved the final manuscript.

REFERENCES

- Åttman, E., Korhonen, P., Tammela, O., Vuento, R., Aittoniemi, J., Syrjänen, J., et al. (2018). A Serratia marcescens outbreak in a neonatal intensive care unit was successfully managed by rapid hospital hygiene interventions and screening. Acta Paediatr. 107, 425–429. doi: 10.1111/apa.14132
- Agirre, J., Aloria, K., Arizmendi, J. M., Iloro, I., Elortza, F., Sánchez-Eugenia, R., et al. (2011). Capsid protein identification and analysis of mature Triatoma virus (TrV) virions and naturally occurring empty particles. *Virology* 409, 91–101. doi: 10.1016/j.virol.2010.09.034
- Alevi, K. C., Moreira, F. F., Jurberg, J., and Azeredo-Oliveira, M. T. (2016). Description of the diploid chromosome set of *Triatoma pintodiasi* (Hemiptera, Triatominae). *Genet. Mol. Res.* 15:gmr.15026343. doi: 10.4238/gmr.15026343
- Alevi, K. C., Rosa, J. A., and Azeredo-Oliveira, M. T. (2014). Cytotaxonomy of the Brasiliensis subcomplex and the *Triatoma brasiliensis* complex (Hemiptera: Reduviidae: Triatominae). *Zootaxa* 3838, 583–589. doi: 10.11646/zootaxa.3838.5.7
- Almeida, C. E., Folly-Ramos, E., Peterson, A. T., Lima-Neiva, V., Gumiel, M., Duarte, R., et al. (2009). Could the bug *Triatoma sherlocki* be vectoring Chagas disease in small mining communities in Bahia, Brazil? *Med. Vet. Entomol.* 23, 410–417. doi: 10.1111/j.1365-2915.2009.00822.x
- Azambuja, P., Feder, D., and Garcia, E. S. (2004). Isolation of Serratia marcescens in the midgut of Rhodnius prolixus: impact on the establishment of the parasite Trypanosoma cruzi in the vector. Exp. Parasitol. 107, 89–96. doi: 10.1016/j.exppara.2004.04.007
- Banerjee, R., Banerjee, B. D., Chaudhury, S., and Hati, A. K. (1991). Transmission of viable *Mycobacterium leprae* by *Aedes aegypti* from lepromatous leprosy patients to the skin of mice through interrupted feeding. *Lepr. Rev.* 62, 21–26.
- Benchimol, J. L., and Romero Sa, M. (2003). Adolpho Lutz and controversies over the transmission of leprosy by mosquitoes. *Hist. Cienc Saude Manguinhos* 10(Suppl. 1), 49–93. doi: 10.1590/S0104-59702003000400004
- Billingsley, P. F., and Downe, A. E. (1986). The surface morphology of the midgut cells of *Rhodnius prolixus* Stål (Hemiptera: Reduviidae) during blood digestion. *Acta Trop.* 43, 355–366.
- Caballero, Z. C., Costa-Martins, A. G., Ferreira, R. C., P. Alves, J. M., and Serrano, M. G., Camargo, E. P., et al. (2015). Phylogenetic and syntenic data support a single horizontal transference to a Trypanosoma ancestor of a prokaryotic proline racemase implicated in parasite evasion from host defences. *Parasit. Vectors* 8:222. doi: 10.1186/s13071-015-0829-y
- Calzada, J. E., Pineda, V., Garisto, J. D., Samudio, F., Santamaria, A. M., and Saldaña, A. (2010). Human trypanosomiasis in the eastern region of the Panama Province: new endemic areas for Chagas disease. *Am. J. Trop. Med. Hyg.* 82, 580–582. doi: 10.4269/ajtmh.2010.09-0397

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fcimb. 2018.00405/full#supplementary-material

- Candeias, J. A. N., Forattini, O. P., and Vieira, J. G. (1976). Hepatitis B antigen (HBsAg) in wild caught Triatominae in Brazil. *Revista de Saúde Pública* 10, 267–268. doi: 10.1590/S0034-89101976000300007
- Carvalho, R. G. C., do Rosário Souza Carneiro, I. C., Pinheiro, M. S., da Costa Pinheiro, S., Roffe Azevedo, P. S., dos Santos, S. D., et al. (2010). Phenotypic and genotypic characterization of *Serratia marcescens* from a Neonatal Unit in Belém, Pará State, Brazil. *Revista Pan Amazônica de Saúde* 1, 101–106. doi: 10.5123/S2176-62232010000100015
- Castro, G. V. S., Ribeiro, M. A. L., Ramos, L. J., Oliveira, J., Rosa, J. A. D., Camargo, L. M. A., et al. (2017). Rhodnius stali: new vector infected by *Trypanosoma rangeli* (Kinetoplastida, Trypanosomatidae). *Rev. Soc. Brasil. Med. Trop.* 50, 829–832. doi: 10.1590/0037-8682-0054-2017
- Castro, M. C., Barrett, T. V., Santos, W. S., Abad-Franch, F., and Rafael, J. A. (2010). Attraction of Chagas disease vectors (Triatominae) to artificial light sources in the canopy of primary Amazon rainforest. *Mem. Inst. Oswaldo Cruz* 105, 1061–1064. doi: 10.1590/S0074-02762010000800019
- Cavalier-Smith, T. (2016). Higher classification and phylogeny of Euglenozoa. *Eur. J. Protistol.* 56, 250–276. doi: 10.1016/j.ejop.2016.09.003
- Cavassin, F. B., Kuehn, C. C., Kopp, R. L., Thomaz-Soccol, V., Da Rosa, J. A., Luz, E., et al. (2014). Genetic variability and geographical diversity of the main Chagas' disease vector *Panstrongylus megistus* (Hemiptera: Triatominae) in Brazil based on ribosomal DNA intergenic sequences. *J. Med. Entomol.* 51, 616–628. doi: 10.1603/ME13073
- Coura, J. R., Fernandes, O., Arboleda, M., Barrett, T. V., Carrara, N., Degrave, W., et al. (1996). Human infection by *Trypanosoma rangeli* in the Brazilian Amazon. *Trans. R. Soc. Trop. Med. Hygiene* 90, 278–279.
- Cuba Cuba, A. (1998). Review of biological and diagnostic aspects of *Trypanosoma* (Herpetosoma) *rangeli. Revista Soc. Brasil. Med. Tropical* 31, 207–220.
- Cullen, M. M., Trail, A., Robinson, M., Keaney, M., and Chadwick, P. R. (2005). Serratia marcescens outbreak in a neonatal intensive care unit prompting review of decontamination of laryngoscopes. J. Hosp. Infect. 59, 68–70. doi: 10.1016/j.jhin.2004.08.003
- Curtis-Robles, R., Hamer, S. A., Lane, S., Levy, M. Z., and Hamer, G. L. (2018). Bionomics and spatial distribution of triatomine vectors of. Am. J. Trop. Med. Hyg. 98, 113–121. doi: 10.4269/ajtmh.17-0526
- Czibener, C., La Torre, J. L., Muscio, O. A., Ugalde, R. A., and Scodeller, E. A. (2000). Nucleotide sequence analysis of Triatoma virus shows that it is a member of a novel group of insect RNA viruses. *J. Gen. Virol.* 81(Pt 4), 1149–1154. doi: 10.1099/0022-1317-81-4-1149
- da Rosa, J. A., Justino H. H. G., Nascimento J. D., Mendonça V. J., Rocha, C. S., de Carvalho D. B., et al. (2017). A new species of *Rhodnius* from Brazil (Hemiptera, Reduviidae, Triatominae). *Zookeys* 675, 1–25. doi: 10.3897/zookeys.675. 12024

- Da Silva, F. M., Marcili, A., Lima, L., Cavazzana, M. Jr., Ortiz, P. A., Campaner, M., et al. (2009). *Trypanosoma rangeli* isolates of bats from Central Brazil: genotyping and phylogenetic analysis enable description of a new lineage using spliced-leader gene sequences. *Acta Trop.* 109, 199–207. doi: 10.1016/j.actatropica.2008.11.005
- Da Silva, F. M., Noyes, H., Campaner, M., Junqueira, A. C., Coura, J. R., Añez, N., et al. (2004). Phylogeny, taxonomy and grouping of *Trypanosoma rangeli* isolates from man, triatomines and sylvatic mammals from widespread geographical origin based on SSU and ITS ribosomal sequences. *Parasitology* 129, 549–561. doi: 10.1017/S0031182004005931
- D'Alessandro, A. (1976). "Biology of Trypanosoma (Herpetosoma) rangeli Tejera, (1920)," in *Biology of the Kinetoplastida*, Vol. 1, eds W. Lumsden and D. Evans (London: Academic Press), 327–403.
- Dario, M. A., Lisboa, C. V., Costa, L. M., Moratelli, R., Nascimento, M. P., Costa, L. P., et al. (2017). High *Trypanosoma* spp. diversity is maintained by bats and triatomines in Espírito Santo state, Brazil. *PLoS ONE* 12:e0188412. doi: 10.1371/journal.pone.0188412
- Dario, M. A., Rodrigues, M. S., Barros, J. H., Xavier, S. C., D'Andrea, P. S., Roque, A. L., et al. (2016). Ecological scenario and *Trypanosoma cruzi* DTU characterization of a fatal acute Chagas disease case transmitted orally (Espírito Santo state, Brazil). *Parasit. Vectors* 9:477. doi: 10.1186/s13071-016-1754-4
- de Moraes, M. H., Guarneri, A. A., Girardi, F. P., Rodrigues, J. B., Eger, I., Tyler, K. M., et al. (2008). Different serological cross-reactivity of *Trypanosoma rangeli* forms in *Trypanosoma cruzi*-infected patients sera. *Parasit. Vectors* 1:20. doi: 10.1186/1756-3305-1-20
- de Scorza, C., Urdaneta-Morales, S., and Tejero, F. (1986). Trypanosoma (Herpetosoma) rangeli Tejera, 1920: Preliminary report on histopathology in experimentally infected mice. *Revista do Instituto de Medicina Tropical de São Paulo* 28, 371–378.
- de Souza-Araujo, H.C. (1942). Poderá o carrapato transmitir a lepra? *Memórias do Instituto Oswaldo Cruz* 37:95.
- Desikan, K. V., and Sreevatsa. (1995). Extended studies on the viability of Mycobacterium leprae outside the human body. Lepr. Rev. 66, 287–295.
- Desquesnes, M., Bosseno, M. F., and Brenière, S. F. (2007). Detection of Chagas infections using Trypanosoma evansi crude antigen demonstrates high cross-reactions with *Trypanosoma cruzi*. *Infect. Genet. Evol.* 7, 457–462. doi: 10.1016/j.meegid.2007.01.007.
- Dias, J. C. P., and Schofield, C. J. (1998). Controle da transmissão transfusional da doença de Chagas na Iniciativa do Cone Sul. *Revista da Sociedade Brasileira de Medicina Tropical* 31, 373–383.
- Domier, L. L., McCoppin, N. K., and D'Arcy, C. J. (2000). Sequence requirements for translation initiation of Rhopalosiphum padi virus ORF2. *Virology* 268, 264–271. doi: 10.1006/viro.2000.0189
- Douglas, A. E. (2015). Multiorganismal insects: diversity and function of resident microorganisms. Annu. Rev. Entomol. 60, 17–34. doi: 10.1146/annurev-ento-010814-020822
- Dujardin, J. P., Lam, T. X., Khoa, P. T., and Schofield, C. J. (2015). The rising importance of Triatoma rubrofasciata. *Mem. Inst. Oswaldo Cruz* 110, 319–323. doi: 10.1590/0074-02760140446
- Edwards, M. S., Kelly, K. S., and Susan, P. M. (2017). Addressing the challenges of chagas disease: an emerging health concern in the United States. *Infect. Dis. Clin. Pract.* 25, 118–125. doi: 10.1097/IPC.000000000000512
- Espinosa-Álvarez, O., Ortiz, P. A., Lima, L., Costa-Martins, A. G., Serrano, M. G., Herder, S., et al. (2018). *Trypanosoma rangeli* is phylogenetically closer to Old World trypanosomes than to *Trypanosoma cruzi*. *Int. J. Parasitol.* 48, 569–584. doi: 10.1016/j.ijpara.2017.12.008
- Fellet, M. R., Lorenzo, M. G., Elliot, S. L., Carrasco, D., and Guarneri, A. A. (2014). Effects of infection by *Trypanosoma cruzi* and *Trypanosoma rangeli* on the reproductive performance of the vector *Rhodnius prolixus*. *PLoS ONE* 9:e105255. doi: 10.1371/journal.pone.0105255
- Ferreira Lde, L., Pereira, M. H., and Guarneri, A. A. (2015). Revisiting *Trypanosoma rangeli* transmission involving susceptible and non-susceptible hosts. *PLoS ONE* 10:e0140575. doi: 10.1371/journal.pone.0140575
- Ferreira, L. L., Lorenzo, M. G., Elliot, S. L., and Guarneri, A. A. (2010). A standardizable protocol for infection of *Rhodnius prolixus* with *Trypanosoma rangeli*, which mimics natural infections and reveals physiological effects of infection upon the insect. *J. Invertebr. Pathol.* 105, 91–97. doi: 10.1016/j.jip.2010.05.013

- Figueiredo, A. R. (1995). Isolamento da Microbiota Bacteriana de Triatomíneos e Persistência do Enterobacter cloacae em Rhodnius prolixus Stal (1859). MSc. Thesis, Instituto Oswaldo Cruz.
- Figueiredo, R. C., Rosa, D. S., and Soares, M. J. (2000). Differentiation of *Trypanosoma cruzi* epimastigotes: metacyclogenesis and adhesion to substrate are triggered by nutritional stress. *J. Parasitol.* 86, 1213–1218. doi: 10.1645/ 0022-3395(2000)086[1213:DOTCEM]2.0.CO;2
- Fleisch, F., Zimmermann-Baer, U., Zbinden, R., Bischoff, G., Arlettaz, R., Waldvogel, K., et al. (2002). Three consecutive outbreaks of Serratia marcescens in a neonatal intensive care unit. *Clin. Infect. Dis.* 34, 767–773. doi: 10.1086/339046
- Frankel, A. D., and Young, J. A. (1998). HIV-1: fifteen proteins and an RNA. Annu. Rev. Biochem. 67, 1–25. doi: 10.1146/annurev.biochem.67.1.1
- Garcia, E. S., Genta, F. A., de Azambuja, P., and Schaub, G. A. (2010). Interactions between intestinal compounds of triatomines and *Trypanosoma cruzi*. *Trends Parasitol.*, 26, 499–505. doi: 10.1016/j.pt.2010.07.003
- Garcia, E. S., Ratcliffe, N. A., Whitten, M. M., Gonzalez, M. S., and Azambuja, P. (2007). Exploring the role of insect host factors in the dynamics of *Trypanosoma cruzi-Rhodnius prolixus* interactions. J. Insect Physiol. 53, 11–21. doi: 10.1016/j.jinsphys.2006.10.006
- Gil-Jaramillo, N., Motta, F. N., Favali, C. B., Bastos, I. M., and Santana, J. M. (2016). Dendritic cells: a double-edged sword in immune responses during chagas disease. *Front. Microbiol.* 7:1076. doi: 10.3389/fmicb.2016.01076
- González-Juarbe, N., Mares, C. A., Hinojosa, C. A., Medina, J. L., Cantwell, A., Dube, P. H., et al. (2015). Requirement for Serratia marcescens cytolysin in a murine model of hemorrhagic pneumonia. *Infect. Immun.* 83, 614–624. doi: 10.1128/IAI.01822-14
- Granato, Hernandes, C. F., de Mendonça, Silva Pinto, P. L., Neto, V. A., Veronesi, R., and Tolezano, E. (1987). Papel de triatomíneos na transmissão de infecção pelo vírus da hepatite tipo B, em diferentes normas clínicas da doença. *Rev. Hosp. Clin. Fac. Med. Sao Paulo* 42, 173–175.
- Gransden, W. R., Webster, M., French, G. L., and Phillips, I. (1986). An outbreak of Serratia marcescens transmitted by contaminated breast pumps in a special care baby unit. J. Hosp. Infect. 7, 149–154.
- Grimont, P. A. D., and Grimont, F. (1978). The genus Serratia. Ann. Rev. Microbiol. 32, 221–248.
- Grimont, P. A. D., Grimont, F., and Lysenko, O. (1979). Species and biotype identification of Serratia strains associated with insects. *Curr. Microbiol.* 2, 139–142.
- Grisard, E. C., Steindel, M., Guarneri, A. A., Eger-Mangrich, I., Campbell, D. A., and Romanha, A. J. (1999a). Characterization of *Trypanosoma rangeli* strains isolated in Central and South America: an overview. *Mem. Inst. Oswaldo Cruz* 94, 203–209.
- Grisard, E. C., Steindel, M., Guarneri, A. A., Eger-Mangrich, I., Campbell, D. A., and Romanha, A. J. (1999b). Characterization of *Trypanosoma rangeli* strains isolated in Central and South America: an overview. *Memórias do Instituto Oswaldo Cruz* 94, 203–209.
- Guarneri, A. A., Pereira, M. H., and Diotaiuti, L. (2000). Influence of the blood meal source on the development of *Triatoma infestans*, *Triatoma brasiliensis*, *Triatoma sordida*, and *Triatoma pseudomaculata* (Heteroptera, Reduviidae). J. Med. Entomol. 37, 373–379. doi: 10.1093/jmedent/37.3.373
- Guhl, F., Hudson, L., Marinkelle, C. J., Jaramillo, C. A., and Bridge, D. (1987). Clinical *Trypanosoma rangeli* infection as a complication of Chagas 'disease. *Parasitology* 94, 475–484.
- Guhl, F., and Marinkelle, C. J. (1982). Antibodies against *Trypanosoma cruzi* in mice infected with *T. rangeli. Ann. Trop. Med. Parasitol.* 76:361.
- Guhl, F., and Vallejo, G. A. (2003). Trypanosoma (Herpetosoma) rangeli Tejera, 1920: an updated review. Mem. Inst. Oswaldo Cruz 98, 435–442. doi: 10.1590/S0074-02762003000400001
- Higgs, S., and Beaty, B. J. (2004). "Natural cycles of vector-borne pathogens," in *The Biology of Disease Vectors*, ed W. H. Marquardt (Burlington, NJ: Elsevier Academic Press), 167–186.
- Hoare, C. A. (1972). The trypanosomes of mammals. A Zoological Monograph. Oxford; Edinburgh: Blackwell Scientific Publication.
- Jang, T. N., Fung, C. P., Yang, T. L., Shen, S. H., Huang, C. S., and Lee, S. H. (2001). Use of pulsed-field gel electrophoresis to investigate an outbreak of Serratia marcescens infection in a neonatal intensive care unit. *J. Hosp. Infect.* 48, 13–19. doi: 10.1053/jhin.2001.0947

- Johnson, K. N., and Christian, P. D. (1998). The novel genome organization of the insect picorna-like virus Drosophila C virus suggests this virus belongs to a previously undescribed virus family. J. Gen. Virol. 79 (Pt 1), 191–203. doi: 10.1099/0022-1317-79-1-191
- Jones, B. L., Gorman, L. J., Simpson, J., Curran, E. T., McNamee, S., Lucas, C., et al. (2000). An outbreak of Serratia marcescens in two neonatal intensive care units. *J. Hosp. Infect.* 46, 314–319. doi: 10.1053/jhin.2000.0837
- Jurberg, J., Rodrigues, J. M. S., Moreira, F. F. F., Dale, C., Cordeiro, I. R. S., and Valdir, D., Jr. (2014). Atlas Iconográfico dos triatomíneos do Brasil (Vetores da doença de Chagas). Laboratorio Nacional e Internacional de Referencia em Taxonomia de Triatomineos, Instituto Oswaldo Cruz.
- Kirchheimer, W. F. (1976). The role of arthropods in the transmission of leprosy. Int. J. Lepr. Other Mycobact. Dis. 44, 104–107.
- Klotz, J. H., Dorn, P. L., Logan, J. L., Stevens, L., Pinnas, J. L., Schmidt, J. O., et al. (2010). "Kissing bugs": potential disease vectors and cause of anaphylaxis. *Clin. Infect. Dis.* 50, 1629–1634. doi: 10.1086/652769
- Kollien, A. H., and Schaub, G. A. (2000). The development of *Trypanosoma cruzi* in triatominae. *Parasitol. Today* 16, 381–387. doi: 10.1016/S0169-4758(00)01724-5
- Lainson, R., Shaw, J. J., Fraiha, H., Miles, M. A., and Draper, C. C. (1979). Chagas's Disease in the Amazon Basin: 1. *Trypanosoma cruzi* infections in silvatic mammals, triatomine bugs and man in the State of Pará, North Brazil. *Trans. R. Soc. Trop. Med. Hyg.* 73, 193–204.
- Laroche, M., Berenger, J. M., Mediannikov, O., Raoult, D., and Parola, P. (2017). Detection of a potential new Bartonella Species Candidatus Bartonella rondoniensis in human biting kissing bugs (Reduviidae; Triatominae). *PLoS Negl. Trop. Dis.* 11:e0005297. doi: 10.1371/journal.pntd.0005297
- Lazzari, C. R., Pereira, M. H., and Lorenzo, M. G. (2013). Behavioural biology of Chagas disease vectors. *Mem. Inst. Oswaldo Cruz* 108(Suppl. 1), 34–47. doi: 10.1590/0074-0276130409
- Lent, H., and Wygodzinsky, P. (1979). Revision of the Triatominae (Hemiptera, Reduviidae) and their significanse as vectors of Chagas disease. Bull. Am. Mus. Nat. Hist. 163, 125–520.
- Liang, T. J. (2009). Hepatitis B: the virus and disease. *Hepatology* 49(5 Suppl), S13-21. doi: 10.1002/hep.22881
- Maguiña, C., and Gotuzzo, E. (2000). Bartonellosis. New and old. *Infect. Dis. Clin.* North Am. 14, 1–22. doi: 10.1016/S0891-5520(05)70215-4
- Mahlen, S. D. (2011). Serratia infections: from military experiments to current practice. Clin. Microbiol. Rev. 24, 755–791. doi: 10.1128/CMR.00017-11
- Maia Da Silva, F., Junqueira, A. C., Campaner, M., Rodrigues, A. C., Crisante, G., Ramirez, L. E., et al. (2007). Comparative phylogeography of *Trypanosoma rangeli* and Rhodnius (Hemiptera: Reduviidae) supports a long coexistence of parasite lineages and their sympatric vectors. *Mol. Ecol.* 16, 3361–3373. doi: 10.1111/j.1365-294X.2007.03371.x
- Maia da Silva, F., Rodrigues, A. C., Campaner, M., Takata, C. S., Brigido, M. C., Junqueira, A. C., et al. (2004). Randomly amplified polymorphic DNA analysis of *Trypanosoma rangeli* and allied species from human, monkeys and other sylvatic mammals of the Brazilian Amazon disclosed a new group and a species-specific marker. *Parasitology* 128(Pt 3), 283–294. doi: 10.1017/S0031182003004554
- Maillard, R., Petit, E., Chomel, B., Lacroux, C., Schelcher, F., Vayssier-Taussat, M., et al. (2007). Endocarditis in cattle caused by *Bartonella bovis. Emerg. Infect. Dis.* 13, 1383–1385. doi: 10.3201/eid1309.070236
- Maragakis, L. L., Winkler, A., Tucker, M. G., Cosgrove, S. E., Ross, T., Lawson, E., et al. (2008). Outbreak of multidrug-resistant Serratia marcescens infection in a neonatal intensive care unit. *Infect. Control Hosp. Epidemiol.* 29, 418–423. doi: 10.1086/587969
- Marliére, N. P., Latorre-Estivalis, J. M., Lorenzo, M. G., Carrasco, D., Alves-Silva, J., Rodrigues, Jde, O., et al. (2015). Trypanosomes modify the behavior of their insect hosts: effects on locomotion and on the expression of a related Gene. *PLoS Negl. Trop. Dis.* 9:e0003973. doi: 10.1371/journal.pntd.0003973
- Marti, G. A., Echeverria, M. G., Susevich, M. L., Becnel, J. J., Pelizza, S. A., and García, J. J. (2009). Prevalence and distribution of parasites and pathogens of triatominae from Argentina, with emphasis on *Triatoma infestans* and Triatoma virus TrV. J. Invertebr. Pathol. 102, 233–237. doi: 10.1016/j.jip.2009.06.010
- Marti, G. A., Ragone, P., Balsalobre, A., Ceccarelli, S., Susevich, M. L., Diosque, P., et al. (2017). Can Triatoma virus inhibit infection of *Trypanosoma cruzi*

(Chagas, 1909) in *Triatoma infestans* (Klug)? A cross infection and coinfection study. *J. Invertebr. Pathol.* 150, 101–105. doi: 10.1016/j.jip.2017. 09.014

- Matsuoka, M., Izumi, S., Budiawan, T., Nakata, N., and Saeki, K. (1999). *Mycobacterium leprae* DNA in daily using water as a possible source of leprosy infection. *Indian J. Lepr.* 71, 61–67.
- McFadzean, J. A., and Macdonald, W. W. (1961). An investigation of the possible role of mosquitoes and bed bugs in the transmission of leprosy. *Trans. R. Soc. Trop. Med. Hyg.* 55, 232–234.
- Minnick, M. F., Anderson, B. E., Lima, A., Battisti, J. M., Lawyer, P. G., and Birtles, R. J. (2014). Oroya fever and verruga peruana: bartonelloses unique to South America. *PLoS Negl. Trop. Dis.* 8:e2919. doi: 10.1371/journal.pntd.0002919
- Montagnani, C., Cocchi, P., Lega, L., Campana, S., Biermann, K. P., Braggion, C., et al. (2015). Serratia marcescens outbreak in a neonatal intensive care unit: crucial role of implementing hand hygiene among external consultants. *BMC Infect. Dis.* 15:11. doi: 10.1186/s12879-014-0734-6
- Montgomery, S. P., Starr, M. C., Cantey, P. T., Edwards, M. S., and Meymandi, S.
 K. (2014). Neglected parasitic infections in the United States: Chagas disease.
 Am. J. Trop. Med. Hyg. 90, 814–818. doi: 10.4269/ajtmh.13-0726
- Mühldorfer, K. (2013). Bats and bacterial pathogens: a review. Zoonoses Public Health 60, 93–103. doi: 10.1111/j.1863-2378.2012.01536.x
- Mullins, K. E., Hang, J., Jiang, J., Leguia, M., Kasper, M. R., Ventosilla, P., et al. (2015). Description of *Bartonella ancashensis* sp. nov., isolated from the blood of two patients with verruga peruana. *Int. J. Syst. Evol. Microbiol.* 65, 3339–3343. doi: 10.1099/ijsem.0.000416
- Muscio, O., Bonder, M. A., La Torre, J. L., and Scodeller, E. A. (2000). Horizontal transmission of triatoma virus through the fecal-oral route in *Triatoma infestans* (Hemiptera: Triatomidae). J. Med. Entomol. 37, 271–275. doi: 10.1603/0022-2585-37.2.271
- Muscio, O. A., La Torre, J., Bonder, M. A., and Scodeller, E. A. (1997). Triatoma virus pathogenicity in laboratory colonies of *Triatoma infestans* (Hemiptera:Reduviidae). J. Med. Entomol. 34, 253–256.
- Muscio, O. A., La Torre, J. L., and Scodeller, E. A. (1988). Characterization of Triatoma virus, a picorna-like virus isolated from the triatomine bug *Triatoma infestans. J. Gen. Virol.* 69(Pt 11), 2929–2934. doi: 10.1099/0022-1317-69-11-2929
- Muscio, O. A., LaTorre, J. L., and Scodeller, E. A. (1987). Small nonoccluded viruses from triatomine bug *Triatoma infestans* (Hemiptera: Reduviidae). *J. Invertebr. Pathol.* 49, 218–220.
- Neumann Ada, S., Dias Fde, A., Ferreira Jda, S., Fontes, A. N., Rosa, P. S., Macedo, R. E., et al. (2016). Experimental infection of *Rhodnius prolixus* (Hemiptera, Triatominae) with *Mycobacterium leprae* indicates potential for leprosy transmission. *PLoS ONE* 11:e0156037. doi: 10.1371/journal.pone.01 56037
- Nuzzo, S., Amato Neto, V., Braz, L., Silva, M., Oliveira, M., Castilho, M., et al. (1998). Evaluation of presence of protein 24 from HIV in feces of *Triatoma infestans* fed blood from HIV positive patients. *Revista de Saúde Pública* 32, 464–466. doi: 10.1590/S0034-891019980005 00009
- Ocaña-Mayorga, S., Aguirre-Villacis, F., Pinto, C. M., Vallejo, G. A., and Grijalva, M. J. (2015). Prevalence, genetic characterization, and 18S small subunit ribosomal RNA diversity of *Trypanosoma rangeli* in triatomine and mammal hosts in endemic areas for Chagas disease in Ecuador. *Vector Borne Zoonot. Dis.* 15, 732–742. doi: 10.1089/vbz.2015.1794
- Ortiz, P. A., Maia da Silva, F., Cortez, A. P., Lima, L., Campaner, M., Pral, E. M., et al. (2009). Genes of cathepsin L-like proteases in *Trypanosoma rangeli* isolates: markers for diagnosis, genotyping and phylogenetic relationships. *Acta Trop.* 112, 249–259. doi: 10.1016/j.actatropica.2009. 07.036
- Osorio, Y., Travi, B. L., Palma, G. I., and Saravia, N. G. (1995). Infectivity of *Trypanosoma rangeli* in a promonocytic mammalian cell line. *J. Parasitol.* 81, 687–693.
- Otálora-Luna, F., A., Pérez-Sánchez, J., Sandoval, C., and Aldana, E. (2015). Evolution of hematophagous habit in Triatominae (Heteroptera: Reduviidae). *Rev. Chil. de Hist. Nat.* 88:4. doi: 10.1186/s40693-014-0032-0
- Paige, C. F., Scholl, D. T., and Truman, R. W. (2002). Prevalence and incidence density of *Mycobacterium leprae* and *Trypanosoma cruzi* infections within

a population of wild nine-banded armadillos. Am. J. Trop. Med. Hyg. 67, 528-532. doi: 10.4269/ajtmh.2002.67.528

- Paim, R. M. M., Pereira, M. H., Araújo, R. N., Gontijo, N. F., and Guarneri, A. A. (2013). The interaction between *Trypanosoma rangeli* and the nitrophorins in the salivary glands of the triatomine *Rhodnius prolixus* (Hemiptera; Reduviidae). *Insect Biochem. Mol. Biol.* 43, 229–236. doi: 10.1016/j.ibmb.2012.12.011
- Peterson, J. K., and Graham, A. L. (2016). What is the 'true' effect of *Trypanosoma rangeli* on its triatomine bug vector? *J. Vector Ecol.* 41, 27–33. doi: 10.1111/jvec.12190
- Peterson, J. K., Graham, A. L., Elliott, R. J., Dobson, A. P., and Triana Chávez, O. (2016). *Trypanosoma cruzi-Trypanosoma rangeli* coinfection ameliorates negative effects of single trypanosome infections in experimentally infected *Rhodnius prolixus*. *Parasitology* 143, 1157–1167. doi: 10.1017/S0031182016000615
- Pinto, C. M., Kalko, E. K., Cottontail, I., Wellinghausen, N., and Cottontail, V. M. (2012). TcBat a bat-exclusive lineage of *Trypanosoma cruzi* in the Panama Canal Zone, with comments on its classification and the use of the 18S rRNA gene for lineage identification. *Infect. Genet. Evol.* 12, 1328–1332. doi: 10.1016/j.meegid.2012.04.013
- Pinto, C. M., Ocaña-Mayorga, S., Tapia, E. E., Lobos, S. E., Zurita, A. P., Aguirre-Villacís, F., et al. (2015). Bats, Trypanosomes, and Triatomines in Ecuador: new insights into the diversity, transmission, and origins of *Trypanosoma cruzi* and Chagas disease. *PLoS ONE* 10:e0139999. doi: 10.1371/journal.pone.0139999
- Pons, M. J., Gomes, C., Del Valle-Mendoza, and Ruiz, J. (2016). Carrion's disease: more than a sand fly-vectored illness. *PLoS Pathog.* 12:e1005863. doi: 10.1371/journal.ppat.1005863
- Querido, J. F., Agirre, J., Marti, G. A., Guérin, D. M., and Silva, M. S. (2013). Inoculation of Triatoma virus (Dicistroviridae: Cripavirus) elicits a non-infective immune response in mice. *Parasit. Vectors* 6:66. doi: 10.1186/1756-3305-6-66
- Querido, J. F., Echeverría, M. G., Marti, G. A., Costa, R. M., Susevich, M. L., Rabinovich, J. E., et al. (2015). Seroprevalence of Triatoma virus (Dicistroviridae: Cripaviridae) antibodies in Chagas disease patients. *Parasit. Vectors* 8:29. doi: 10.1186/s13071-015-0632-9
- Rassi, A. Jr., Rassi, A., and Marin-Neto, J. A. (2010). Chagas disease. *Lancet* 375, 1388–1402. doi: 10.1016/S0140-6736(10)60061-X
- Regier, Y., O., Rourke, F., and Kempf, V. A. (2016). Bartonella spp. a chance to establish One Health concepts in veterinary and human medicine. Parasit. Vectors 9:261. doi: 10.1186/s13071-016-1546-x
- Rodríguez-Ruano, S. M., Škochová, V., Rego, R. O. M., Schmidt, J. O., Roachell, W., Hypša, V., et al. (2018). Microbiomes of North American Triatominae: the grounds for Chagas Disease epidemiology. *Front. Microbiol.* 9:1167. doi: 10.3389/fmicb.2018.01167
- Rozas-Dennis, G. S., and Cazzaniga, N. J. (2000). Effects of Triatoma virus (TrV) on the fecundity and moulting of *Triatoma infestans* (Hemiptera: Reduviidae). *Ann. Trop. Med. Parasitol.* 94, 633–641. doi: 10.1080/00034983.2000.118 13586
- Sanchez Clemente, N., Ugarte-Gil, C. A., Solórzano, N., Maguiña, C., Pachas, P., Blazes, D., et al. (2012). Bartonella bacilliformis: a systematic review of the literature to guide the research agenda for elimination. *PLoS Negl. Trop. Dis.* 6:e1819. doi: 10.1371/journal.pntd.0001819
- Sartor, C., Jacomo, V., Duvivier, C., Tissot-Dupont, H., Sambuc, R., and Drancourt, M. (2000). Nosocomial Serratia marcescens infections associated with extrinsic contamination of a liquid nonmedicated soap. *Infect. Control. Hosp. Epidemiol.* 21, 196–199. doi: 10.1086/501743
- Sasaki, J., Nakashima, N., Saito, H., and Noda, H. (1998). An insect picorna-like virus, Plautia stali intestine virus, has genes of capsid proteins in the 3' part of the genome. *Virology* 244, 50–58. doi: 10.1006/viro.1998.9094
- Schaub, G. A. (1989). Does Trypanosoma cruzi stress its vectors? Parasitology Today 5, 185–188.
- Schaub, G. A., Meiser, C. K., and Balczun, C. (2011). "Interactions of *Trypanosoma cruzi* and triatomines," in *Progress in Parasitology*, ed H. Mehlhorn (Berlin; Heidelberg: Springer) 155–178.
- Schofield, C. J. (1988) "The biosystematics of Triatominae," in *Biosystematics of Haematophagous Insects*, Vol. 37, ed M. W. Service (Oxford University Press), 284–312.

- Schuster, J. P., and Schaub, G. A. (2000). *Trypanosoma cruzi*: skin-penetration kinetics of vector-derived metacyclic trypomastigotes. *Int. J. Parasitol.* 30, 1475–1479. doi: 10.1016/S0020-7519(00)00119-3
- Scollard, D. M., Adams, L. B., Gillis, T. P., Krahenbuhl, J. L., Truman, R. W., and Williams, D. L. (2006). The continuing challenges of leprosy. *Clin. Microbiol. Rev.* 19, 338–381. doi: 10.1128/CMR.19.2.338-381.2006
- Sincero, T. C., Stoco, P. H., Steindel, M., Vallejo, G. A., and Grisard, E. C. (2015). *Trypanosoma rangeli* displays a clonal population structure, revealing a subdivision of KP1(-) strains and the ancestry of the Amazonian group. *Int. J. Parasitol.* 45, 225–235. doi: 10.1016/j.ijpara.2014.11.004
- Smith, J. H., Folse, D. S., Long, E. G., Christie, J. D., Crouse, D. T., Tewes, M. E., et al. (1983). Leprosy in wild armadillos (Dasypus novemcinctus) of the Texas Gulf Coast: epidemiology and mycobacteriology. *J. Reticuloendothelial Soc.* 34, 75–88.
- Souza, E. D., Von Atzingen, N. C., Furtado, M. B., de Oliveira, J., Nascimento, J. D., Vendrami, D. P., et al. (2016). Description of *Rhodnius marabaensis* sp. n. (Hemiptera, Reduviidae, Triatominae) from Pará State, Brazil. *ZooKeys* 621, 45–62. doi: 10.3897/zookeys.621.9662
- Tarleton, R. L. (2007). Immune system recognition of *Trypanosoma cruzi*. Curr Opin Immunol 19, 430–434. doi: 10.1016/j.coi.2007.06.003
- Teixeira, A. R., Hecht, M. M., Guimaro, M. C., Sousa, A. O., and Nitz, N. (2011). Pathogenesis of chagas' disease: parasite persistence and autoimmunity. *Clin. Microbiol. Rev.* 24, 592–630. doi: 10.1128/CMR.00063-10
- Truman, R. W., Singh, P., Sharma, R., Busso, P., Rougemont, J., Paniz-Mondolfi, A., et al. (2011). Probable zoonotic leprosy in the southern United States. N. Engl. J. Med. 364, 1626–1633. doi: 10.1056/NEJMoa1010536
- Tsai, Y. L., Chang, C. C., Chuang, S. T., and Chomel, B. B. (2011). Bartonella species and their ectoparasites: selective host adaptation or strain selection between the vector and the mammalian host? *Comp. Immunol. Microbiol. Infect. Dis.* 34, 299–314. doi: 10.1016/j.cimid.2011.04.005
- Turankar, R. P., Lavania, M., Singh, M., Sengupta, U., Siva Sai, K., and Jadhav, R. S. (2016). Presence of viable *Mycobacterium leprae* in environmental specimens around houses of leprosy patients. *Indian J. Med. Microbiol.* 34, 315–321. doi: 10.4103/0255-0857.188322
- Turankar, R. P., Lavania, M., Singh, M., Siva Sai, K. S., and Jadhav, R. S. (2012). Dynamics of *Mycobacterium leprae* transmission in environmental context: deciphering the role of environment as a potential reservoir. *Infect. Genet. Evol.* 12, 121–126. doi: 10.1016/j.meegid.2011.10.023
- Tyler, K. M., and Engman, D. M. (2001). The life cycle of *Trypanosoma cruzi* revisited. *Int. J. Parasitol.* 31, 472–481. doi: 10.1016/S0020-7519(01)00153-9
- Uduman, S. A., Farrukh, A. S., Nath, K. N., Zuhair, M. Y., Ifrah, A., Khawla, A. D., et al. (2002). An outbreak of Serratia marcescens infection in a special-care baby unit of a community hospital in United Arab Emirates: the importance of the air conditioner duct as a nosocomial reservoir. *J. Hosp. Infect.* 52, 175–180. doi: 10.1053/jhin.2002.1308
- Urdaneta-Morales, S., and Tejero, F. (1986). Trypanosoma (Herpetosoma) rangeli Tejera, 1920: intracellular amastigote stages of reproduction in white mice. *Revis. Inst. Med. Trop. São Paulo* 28, 166–169.
- Vallejo, G. A., Guhl, F., and Schaub, G. A. (2009). Triatominae–Trypanosoma cruzi/T. rangeli: vector-parasite interactions. Acta Trop. 110, 137–147. doi: 10.1016/j.actatropica.2008.10.001
- Vallejo, G. A., Suárez, J., Olaya, J. L., Gutierrez, S. A., and Carranza, J. C. (2015). *Trypanosoma rangeli*: un protozoo infectivo y no patógeno para el humano que contribuye al entendimiento de la transmisión vectorial y la infección por *Trypanosoma cruzi*, agente causal de la enfermedad de Chagas. *Revista de la Academia Colombiana de Ciencias Exactas, Físicas y Naturales* 39, 111–122. doi: 10.18257/raccefyn.143
- Villari, P., Crispino, M., Salvadori, A., and Scarcella, A. (2001). Molecular epidemiology of an outbreak of Serratia marcescens in a neonatal intensive care unit. *Infect. Control Hosp. Epidemiol.* 22, 630–634. doi: 10.1086/501834
- Walsh, G. P., Storrs, E. E., Burchfield, H. P., Cotrell, E. H., Vidrine, M. F., and Binford, C. H. (1975). Leprosy-like disease occurring naturally in armadillos. *J. Reticuloendothelial Soc.* 18, 347–351.
- Walter, J., Fletcher, E., Moussaoui, R., Gandhi, K., and Weirauch, C. (2012). Do bites of kissing bugs cause unexplained allergies? Results from a survey in triatomine-exposed and unexposed areas in southern California. *PLoS ONE* 7:e44016. doi: 10.1371/journal.pone.0044016

- WHO (2015). Leprosy Elimination. Available online at: http://www.who.int/lep/ disease/en/ (Accessed March 14, 2018).
- WHO (2016). Global Leprosy Strategy 2016-2020: Accelerating Towards a Leprosy-Free World.
- WHO (2017). Vector-Borne Diseases. Available online at: http://www.who.int/ en/news-room/fact-sheets/detail/vector-borne-diseases (Accessed October 14, 2018).
- WHO (2018a). Chagas Disease (American trypanosomiasis) Epidemiology. Available online at: http://www.who.int/chagas/epidemiology/en/ (Accessed October 10, 2018).
- WHO (2018b). Chagas disease (American trypanosomiasis). Available online at: http://www.who.int/mediacentre/factsheets/fs340/en/ (Accessed July 29, 2018).
- WHO (2018c). HIV/AIDS. Available online at: http://www.who.int/news-room/ fact-sheets/detail/hiv-aids (Accessed September 22, 2018).
- WHO (2018d). *Hepatitis, B.* Available online at: http://www.who.int/news-room/ fact-sheets/detail/hepatitis-b (Accessed October 04, 2018).
- WHO/TDR (2012). Research Priorities for Chagas Disease, Human African Trypanosomiasis and Leishmaniasis. Technical Report of the TDR Disease

Reference Group on Chagas Disease, Human African Trypanosomiasis and *Leishmaniasis*. World Health Organization.

Wilson, J. E., Powell, M. J., Hoover, S. E., and Sarnow, P. (2000). Naturally occurring dicistronic cricket paralysis virus RNA is regulated by two internal ribosome entry sites. *Mol. Cell Biol.* 20, 4990–4999.

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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